

Bacterial Nucleoid: Interplay of DNA Demixing and Supercoiling

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ABSTRACT This work addresses the question of the interplay of DNA demixing and supercoiling in bacterial cells. Demixing of DNA from other globular macromolecules results from the overall repulsion between all components of the system and leads to the formation of the nucleoid, which is the region of the cell that contains the genomic DNA in a rather compact form. Supercoiling describes the coiling of the axis of the DNA double helix to accommodate the torsional stress injected in the molecule by topoisomerases. Supercoiling is able to induce some compaction of the bacterial DNA, although to a lesser extent than demixing. In this work, we investigate the interplay of these two mechanisms with the goal of determining whether the total compaction ratio of the DNA is the mere sum or some more complex function of the compaction ratios due to each mechanism. To this end, we developed a coarse-grained bead-and-spring model and investigated its properties through Brownian dynamics simulations. This work reveals that there actually exist different regimes, depending on the crowder volume ratio and the DNA superhelical density. In particular, a regime in which the effects of DNA demixing and supercoiling on the compaction of the DNA coil simply add up is shown to exist up to moderate values of the superhelical density. In contrast, the mean radius of the DNA coil no longer decreases above this threshold and may even increase again for sufficiently large crowder concentrations. Finally, the model predicts that the DNA coil may depart from the spherical geometry very close to the jamming threshold as a trade-off between the need to minimize both the bending energy of the stiff plectonemes and the volume of the DNA coil to accommodate demixing.

SIGNIFICANCE Many biological processes take place simultaneously in living cells. It is tempting to study each of them separately and rely on the hypothesis that cells behave like the “addition” of the isolated parts. This is, however, not always the case, and this work illustrates this fact. We consider two different processes that are both able to compact the bacterial DNA, namely demixing and supercoiling, and we study how the DNA reacts when subject to both of them simultaneously. Through coarse-grained modeling and Brownian dynamics simulations, we show that the two processes are “additive” only in a limited range of biologically relevant values of the parameters and that their interplay is much more complex outside from this range.

INTRODUCTION

This work addresses the question of the interplay of DNA demixing and supercoiling in bacterial cells. Prokaryotic cells are simpler than eukaryotic ones in many respects, yet several of their fundamental features remain poorly understood. For example, the mechanism leading to the formation of the bacterial nucleoid is a longstanding but still hotly debated question (1–5). The nucleoid is the region of the cell that contains the genomic DNA, together with a certain number of proteins and other macromolecules (6). In contrast with the nucleus of eukaryotic cells, the bacterial

nucleoid is not separated from the rest of the cytosol by a bounding membrane. It nevertheless occupies only a fraction of the cell, whose volume depends sensitively on several factors such as the richness of the nutrients (7–11), the cell cycle step (12,13), and the eventual addition of antibiotics (9–11,14–18). This is quite puzzling because the volume of the unconstrained bacterial genomic DNA in physiological saline conditions (estimated from the worm-like chain model (19)) is approximately 1000 times larger than the volume of the cell. One therefore has to understand why the DNA molecule remains localized inside the nucleoid instead of expanding throughout the cell. It is becoming increasingly clear that the formation of plectonemes, the bridging of DNA duplexes by nucleoid proteins, and the action of short-range attractive forces, which are commonly evoked as the mechanisms responsible for the

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formation of the bacterial nucleoid (1), may not play the leading role in the compaction of the DNA (see, for example, (4) and references therein). In contrast, the 20-year-old proposition that increasing amounts of nonbinding globular macromolecules may be able to compact the genomic DNA gradually (20–24) has recently received strong support, both from the experimental (25–27) and computational (28–33) sides. The proposed mechanism is that the overall repulsion between all components of the system leads to a separation into two phases (34–45), one of them being rich in DNA and poor in the other macromolecule (the nucleoid) and the other one being almost deprived of DNA (the rest of the cytosol). The connectivity of the long DNA molecule and its ability to deform close to and around the proteins to fit in void spaces between proteins play crucial roles in this mechanism in that they induce many-body interaction terms that ultimately result in effective DNA-DNA and protein-protein attraction when DNA is depleted from the regions between particles. Compaction of the genomic DNA through its demixing from other macromolecules of the cytosol is the first point this work focuses on.

The second point deals with supercoiling, that is, the winding about itself of the circular DNA double helix in response to the torsional stress induced by topoisomerases (46). Experimental and theoretical results suggest that supercoiling is able to induce some limited compaction of the bacterial DNA. For example, relaxation of the underwinding of the circular DNA molecule through inhibition of the topoisomerase activity leads to a modest increase in the size of *Escherichia coli* nucleoids (14,47). Moreover, theoretical arguments suggest that the radius of gyration of an unconstrained supercoiled DNA molecule with contour length of 2.6 mm is of the order of 2.5 μm (48), which is smaller than the worm-like chain estimate for a circular chain with the same contour length ($\sim 3.4 \mu\text{m}$) but still significantly larger than the average dimensions of *E. coli* cells.

The question addressed in this study is that of the interplay of these two mechanisms, DNA demixing and supercoiling, which are both able to compact the bacterial DNA, and more precisely, the question of whether the total compaction of the DNA coil is the mere sum or some more complex function of the compaction ratios due to each mechanism. Stated in other words, is the increase of DNA compaction ratio provoked by an increase of crowder density similar for torsionally relaxed (less compact) and supercoiled (more compact) DNA coils? Or, conversely, is the increase of DNA compaction ratio provoked by an increase of torsional stress similar for DNA immersed in a dilute cytosol (less compact DNA coils) and a highly crowded cytosol (more compact DNA coils)? Indeed, plectonemes are composed of two intertwined DNA duplexes and are consequently thicker (estimated diameter in the range 10–32 nm for standard values of the underwinding of in vivo

DNA (48–50) and more rigid (estimated persistence length of ~ 80 nm (48)) than simple duplexes (diameter of ~ 2 nm and persistence length of ~ 50 nm). One may therefore expect that the demixing mechanism is less efficient in compacting plectonemic DNA than linear DNA, which implies that the effects of the two mechanisms do not simply add up.

More generally, understanding the interplay of DNA demixing and supercoiling is important for rationalizing some in vivo observations. For example, the nucleoid of certain bacteria, such as *E. coli*, is divided into different macrodomains (four macrodomains for *E. coli* cells) that display quite different densities of DNA nucleotides (51–54). It is known that certain families of nucleoid proteins probably contribute to the organization of these macrodomains (55–57). On the other hand, it is also known that the DNA molecule is dynamically partitioned into several hundreds of independently supercoiled loops with an average size of ~ 10000 basepairs (bp), which are called topological domains (58,59). One may therefore reason that the difference in DNA density in different macrodomains may result from different levels of supercoiling in the corresponding topological domains, provided that different levels of supercoiling result in different levels of DNA compaction at constant crowder density.

The topology and dynamics of free supercoiled DNA have received much attention from the experimental (47,49,60–64), theoretical (65), and numerical (66–72) points of view. The effects of confinement (73–75) and increasing nucleic acid concentration (72,76) on supercoiled DNA have also been investigated. In contrast, much less work has been devoted to the influence of nonbinding globular macromolecules on the conformations of supercoiled DNA (22,48,77,78). In the perspective of this work, the most striking result is probably the observation that in crowded conditions, the size of supercoiled DNA may exceed that of its linear variant (77), which supports the putative nonadditivity of the two compaction mechanisms. Moreover, condensation experiments suggest that the tight packing of DNA supercoils in condensates is facilitated by the decrease of the diameter of plectonemes rather than by a variation of the writhe/twist ratio (78), which points toward the need for mechanical rearrangements of the plectonemes to accommodate strong compaction ratios. These results were, however, obtained for short plasmids (less than 3000 bp) in free solution and need to be confirmed for longer and confined DNA molecules.

To shed light on the interplay of DNA demixing and supercoiling, a coarse-grained model was developed along the same lines as those used previously to investigate facilitated diffusion (79–81), the interactions of DNA and H-NS nucleoid proteins (82–84), and the formation of the bacterial nucleoid (4,5,32,33,85). Torsional energy was accounted for in the model as described in (86), and the properties of the full model were investigated for different values of the

number of crowders and the superhelical density (i.e., the relative overwinding) of the DNA chain. In particular, the number of crowders was increased up to the jamming threshold, at which strong compaction is known to occur (32,33), and the investigated range of superhelical density values encompasses the estimated value for *E. coli* (87). The results presented in this article reveal that there actually exist different regimes, which are separated by threshold values of the crowder volume ratio and the DNA superhelical density. In particular, a regime in which the effects of DNA demixing and supercoiling on the compaction of the DNA coil simply add up is shown to exist up to moderate values of the superhelical density, whereas the mean radius of the DNA coil ceases to decrease above this threshold and may even increase again for sufficiently large crowder concentrations. Moreover, the model predicts that the DNA coil may depart from the spherical geometry very close to the jamming threshold as a trade-off between the need to minimize the bending energy of stiff plectonemes and the need to minimize the volume of the DNA coil to accommodate demixing.

METHODS

The coarse-grained bead-and-spring model developed for this study is described in detail in Model and Simulations in the Supporting Materials and Methods. In brief, the DNA molecule is modeled, as in (84), as a circular chain of 2880 beads with radius 1.0 nm separated at equilibrium by a distance of 2.5 nm and enclosed in a confinement sphere of radius 120 nm, which represents the cell envelope. Two beads represent 15 bp, and the circular chain therefore corresponds to 21600 bp. Both the contour length of the DNA chain and the volume of the confinement sphere are ~ 200 times smaller than their actual values in *E. coli* cells so that the nucleic acid concentration of the model is close to the in vivo concentration of most bacteria (~ 10 mM). Note that the spherical confinement chamber is adequate for modeling cocci but not bacilli (including *E. coli*), which look rather like capped cylinders. DNA beads interact through stretching, bending, torsional, and electrostatic terms. The bending rigidity constant is chosen so that the model reproduces the known persistence length of double-stranded DNA (~ 50 nm). The torsional energy term is borrowed from (86), and the torsional rigidity is adjusted so that at equilibrium, the writhe contribution accounts for $\sim 70\%$ of the linking number difference (49). The values of the bending and torsional rigidities are close together, in agreement with experimental results (49). Electrostatic repulsion between DNA beads is written as a sum of Debye-Hückel terms, which depend on effective electrostatic charges placed at the center of each bead. The values of these charges are derived from the net linear charge density along a DNA molecule immersed in a buffer with monovalent cations according to Manning's counterion condensation theory (88,89). The value of the Debye length (~ 1 nm) corresponds to a concentration of monovalent salt of 100 mM, which is the value that is generally assumed for the cytoplasm of bacterial cells.

Globular macromolecular crowders are modeled as a variable number N of spheres with radius 7.4 nm and the same electrostatic charge as DNA beads. DNA-crowder and crowder-crowder interactions are expressed as sums of Debye-Hückel potentials so that all components of the systems repel each other. It was shown previously (32,33) that maximal compaction of torsionally relaxed DNA chains is obtained when ρ , the effective crowder volume fraction (Eq. S6), is close to the jamming threshold for hard spheres, that is, $\rho \approx 0.65$.

The properties of this model for supercoiled DNA were investigated by integrating numerically overdamped Langevin equations with time steps of 10 ps for different values of the number of crowders, N , and the superhelical density of the DNA chain, σ . Various sets of trajectories were run with $N = 0, 1500, 1750, 1875,$ and 2000 , which corresponds to effective crowder volume ratios $\rho = 0, 0.49, 0.57, 0.61,$ and 0.65 , respectively. σ was varied from 0 (torsionally relaxed DNA) to -0.08 , thus encompassing the estimated value for *E. coli* cells, $\sigma \approx -0.06$ (87). Simulations were performed by first letting DNA chains with prescribed values of σ equilibrate for 2 ms inside the confinement sphere in the absence of any crowder. The N crowding spheres were then added at random, homogeneously distributed, and nonoverlapping positions, and the complete system was allowed to equilibrate again for 5 ms ($N = 1500$ and 1750), 10 ms ($N = 1875$), or 20 ms ($N = 2000$) to cope with the marked slowing-down of the dynamics close to the jamming threshold. The radius of the DNA coil, R (computed as the mean distance of the DNA beads from the center of mass of the DNA coil), the excess of twist, ΔTw , and the writhe, Wr , were then averaged over time windows of 8 ms ($N = 0$), 15 ms ($N = 1500$ and 1750), 20 ms ($N = 1875$), and up to 60 ms for $N = 2000$. For the sake of better statistics, all results were moreover averaged over four different trajectories with different initial conditions but identical values of N and σ . The error bars shown in the figures correspond to the SD of the four average values obtained from the different trajectories with identical values of N and σ . Temperature T was assumed to be 298 K throughout the study. Representative snapshots extracted from trajectories with $N = 0$ and $N = 1875$ ($\rho = 0.61$) are shown in Figs. 1 and 2, respectively.

RESULTS AND DISCUSSION

The goal of this work is to shed light on the interplay of DNA supercoiling and DNA demixing from other macromolecules with respect to the compaction of the DNA coil. To this end, several sets of trajectories were launched with different values of the effective crowder volume ratio, ρ , and the superhelical density of the DNA, σ . It is

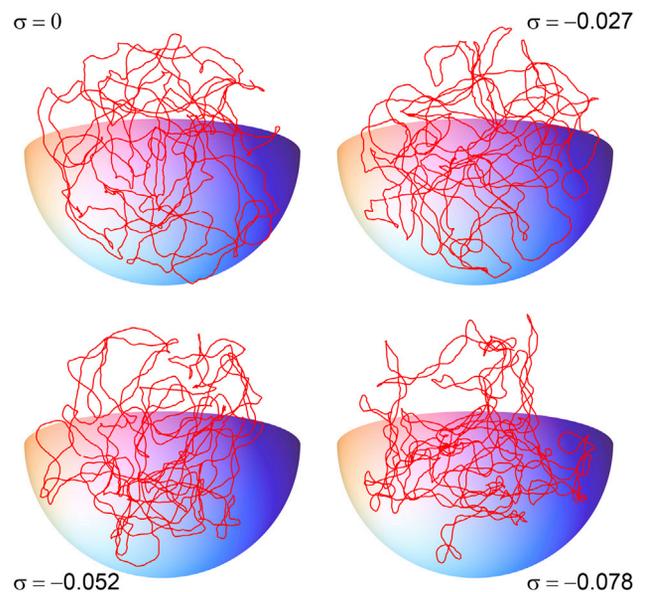


FIGURE 1 Representative snapshots extracted from trajectories with $N = 0$ ($\rho = 0$) and $\sigma = 0, -0.027, -0.052,$ and -0.078 . The solid red line connects the centers of successive DNA beads. Only one-fourth of the confinement sphere is shown. To see this figure in color, go online.

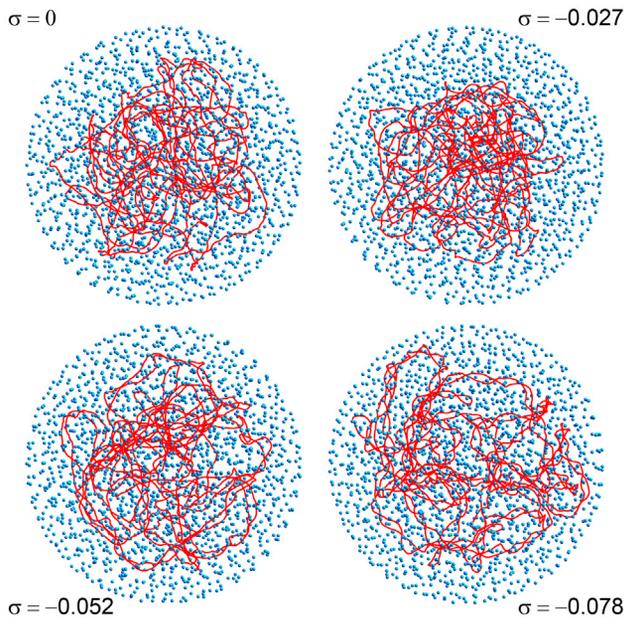


FIGURE 2 Representative snapshots extracted from trajectories with $N = 1875$ ($\rho = 0.61$) and $\sigma = 0, -0.027, -0.052,$ and -0.078 . The solid red line connects the centers of successive DNA beads. Crowding spheres are colored in cyan and represented at $1/5$ of their actual radius so that the DNA chain can be seen through the layers of crowders. The confinement sphere is not shown. To see this figure in color, go online.

reminded that the superhelical density is defined according to $\sigma = \Delta Lk/Lk_0$, where Lk_0 and $Lk_0 + \Delta Lk$ are the linking numbers of the torsionally relaxed and topologically constrained DNA, respectively. The linking number difference ΔLk is the sum of two contributions, namely the excess of twist around the DNA axis, ΔTw , and the writhe, Wr , which quantifies the winding of the DNA axis around itself. The linking number difference $\Delta Lk = \Delta Tw + Wr$ and the superhelical density $\sigma = \Delta Lk/Lk_0$ remain constant as long as the DNA molecule is not nicked. In contrast, the excess of twist ΔTw and the writhe Wr do fluctuate under the influence of thermal noise and external constraints. Note that in this work, ΔTw , Wr , ΔLk , and σ are all negative quantities as a consequence of the underwinding of bacterial DNA and that negative values of ΔTw must be understood as a deficit of twist compared to torsionally relaxed DNA.

The effect on DNA compaction of each mechanism taken separately

The evolution of the compaction of the DNA coil with ρ and σ is summarized in Fig. 3, which shows the evolution of $\langle R \rangle$ with $-\sigma$ for different values of ρ . The blue dashed line with open circular symbols indicates that the mean radius of the DNA coil decreases from 82 nm down to 75 nm when $|\sigma|$ is increased from 0 up to 0.08 in the absence of any crowder ($\rho = 0$). Along this curve, compaction results from the reorganization of the DNA chain, which forms an

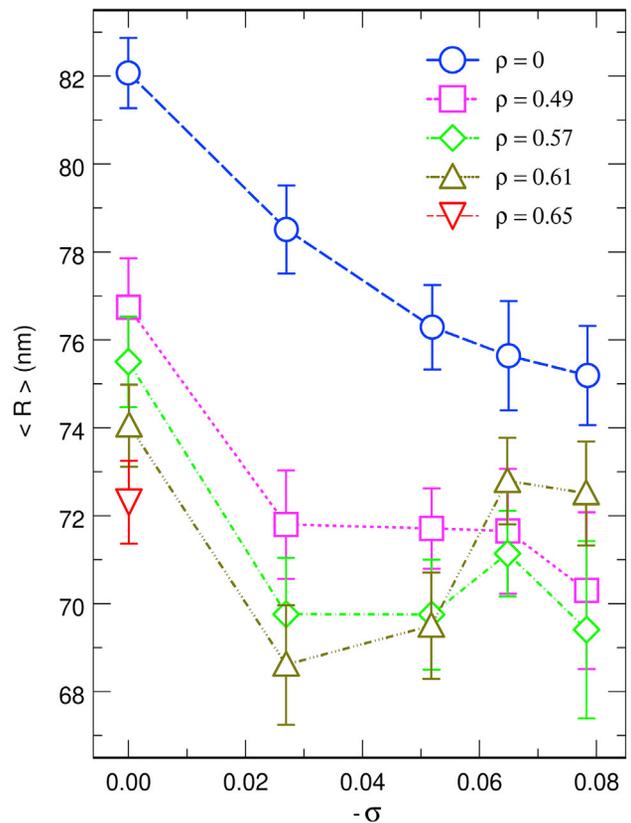


FIGURE 3 Plot of the mean radius of the DNA coil, $\langle R \rangle$, as a function of the opposite of the superhelical density of the DNA chain, $-\sigma$, for values of the effective crowder volume ratio ρ ranging from 0 to 0.65. The lines are guides for the eyes. See text for explanations regarding the points that are missing for $\rho = 0.65$. The error bars correspond to the standard deviation of the average values obtained from 4 different trajectories with identical values of N and σ . To see this figure in color, go online.

increasing number of plectonemes to reduce torsional stress (see Fig. 1). Plectonemes can conveniently be sought for as illustrated in Fig. S1, which displays the index j of the bead located closest to bead i for the equilibrated DNA conformations shown in Fig. 1. The nearest neighbors were searched for with the constraint that $j \notin [i - 10, i + 10]$ so that the algorithm does not systematically select immediate neighbors along the DNA chain. In the vignette for torsionally relaxed DNA ($\sigma = 0$), the points are essentially randomly distributed, except for a discrete accumulation of points close to the diagonal, which correspond to the trivial case in which the search algorithm led to $j = i \pm 11$. In the vignettes for $\sigma = -0.052$ and $\sigma = -0.078$, plectonemes appear instead as well-defined segments parallel to the antidiagonal. Plectonemes also appear in the vignette for $\sigma = -0.027$, but the segments are significantly shorter, thereby pointing toward a poor plectonemic structure. As illustrated in the top plot of Fig. 4 (blue dashed line with open circular symbols) and in agreement with previous work, the mean distance $\langle d \rangle$ between the opposite strands of the plectonemes decreases significantly, like $1/|\sigma|$ (49),

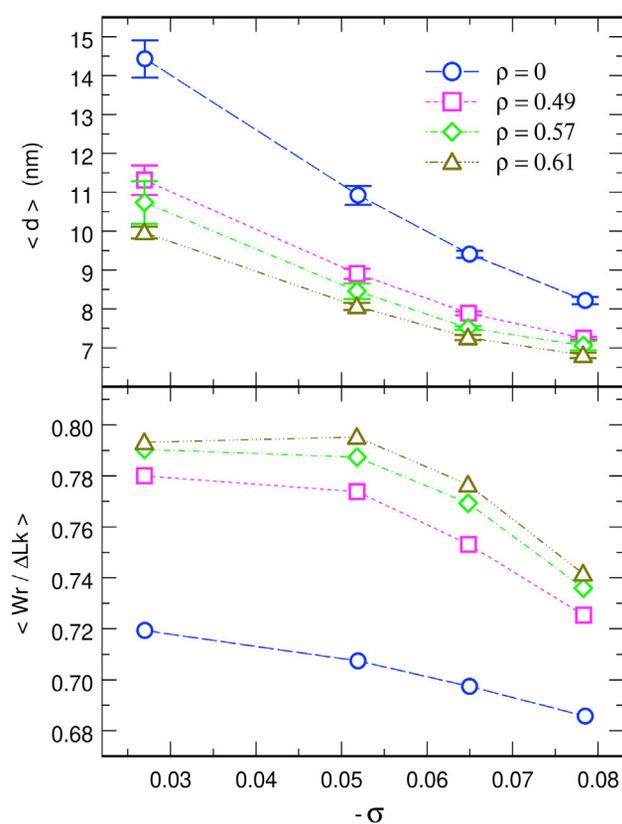


FIGURE 4 Plot as a function of the opposite of the superhelical density of the DNA chain, $-\sigma$, of the mean distance $\langle d \rangle$ between the opposite strands of the plectonemes (*top plot*) and the mean contribution $\langle Wr / \Delta Lk \rangle$ of the writhe to the linking number difference (*bottom plot*), for values of the effective crowder volume ratio ρ ranging from 0 to 0.61. The lines are guides for the eyes. The error bars correspond to the standard deviation of the average values obtained from 4 different trajectories with identical values of N and σ . To see this figure in color, go online.

with increasing values of $|\sigma|$. Moreover, the mean contribution of the writhe to the linking number difference, $\langle Wr / \Delta Lk \rangle$, decreases slightly with increasing values of $|\sigma|$ (*blue dashed line with open circular symbols in the bottom plot of Fig. 4*). It was shown previously that $\langle Wr / \Delta Lk \rangle$ increases with $|\sigma|$ for short DNA sequences (300–3500 bp) and remains nearly constant for a 7 kbp DNA sequence (66). This work therefore confirms that $\langle Wr / \Delta Lk \rangle$ varies in opposite directions as a function of $|\sigma|$ for short (<10 kbp) and long (>10 kbp) DNA sequences.

On the other hand, the points on the ordinate axis ($\sigma = 0$) in Fig. 3 indicate that the mean radius of the torsionally relaxed DNA coil decreases from 82 nm down to 72.3 nm when ρ is increased from 0 to 0.65. Along this axis, compaction of the DNA coil results from the demixing of the DNA chain and the spherical crowders, which are expelled outside from the DNA coil (32,33) (see Fig. 2). As was already observed in previous studies based on somewhat different models (32,33), demixing increases strongly close to the jamming threshold for spherical crowders ($\rho \approx 0.65$). This is more clearly seen in Fig. S2, which shows the evo-

lution of $\langle R \rangle$ with ρ for torsionally relaxed DNA. A further slight increase of ρ beyond 0.65 will probably result in significantly stronger compaction of the DNA coil, as in (32,33), but the dynamics of the system becomes too slow to be numerically tractable with our computer facility.

The purpose of this work is to decipher the rest of Fig. 3, that is, to rationalize the compaction of topologically constrained DNA with increasing crowder volume fraction.

The additive regime at moderate superhelical density

A first remarkable feature of Fig. 3 is that the curves corresponding to different values of ρ are parallel to the curve for $\rho = 0$ up to $-\sigma = 0.027$. This indicates that the effects of the two compaction mechanisms are actually simply additive for such moderate values of the superhelical density. As a consequence, the mean radius of the DNA coil is as small as 68.6 nm for $\rho = 0.61$ and $\sigma = -0.027$. The reason for such additivity is that the plectonemes are still few and loose and do not oppose compaction. This point can be checked in the vignettes for $\sigma = -0.027$ in Figs. 1 and 2, in which plectonemes are hardly noticeable, and Fig. S1, in which the segments of points that signalize plectonemes are significantly shorter than for larger values of $|\sigma|$. Moreover, as was anticipated on the basis of theoretical grounds in (78) and observed upon increasing nucleic acid concentration in (77), the compaction of the DNA coil is facilitated by a decrease of the diameter of the superhelix with increasing crowder concentration. Indeed, the top plot of Fig. 4 indicates that for $\sigma = -0.027$, the mean distance $\langle d \rangle$ between the opposite strands of the plectonemes decreases from 14.4 nm in the absence of any crowder down to 10.0 nm for $\rho = 0.61$. The fact that compaction of the DNA coil upon increase of the concentration of crowders is accompanied by an increase of $\langle Wr / \Delta Lk \rangle$ (see *bottom plot of Fig. 4*) is more surprising because one would rather imagine that compaction is favored by unwinding rather than winding of the supercoils (78). A possible interpretation of this observation is that further winding of the supercoils from $\langle Wr / \Delta Lk \rangle = 0.72$ ($\langle Wr \rangle \approx -40$) to $\langle Wr / \Delta Lk \rangle = 0.79$ ($\langle Wr \rangle \approx -44$) contributes to the reduction of the diameter of the superhelix, which in turn facilitates compaction.

The nonadditive regime at larger superhelical density

Additivity of the effects of DNA supercoiling and demixing on the compaction of DNA coils does however not extend to values of $|\sigma|$ larger than 0.027. This can be checked in Fig. 3, where the curves corresponding to different values of ρ do not remain parallel to the curve for $\rho = 0$ above $|\sigma| = 0.027$. Owing to the uncertainties on the computed values of $\langle R \rangle$ (~ 1 nm), all that can be said safely is

that 1) at moderate crowder volume ratios ($\rho = 0.49$ and $\rho = 0.57$), winding of the plectonemes beyond $|\sigma| = 0.027$ does not result in a significant increase of the compaction of the DNA coil, in contrast with the case of $\rho = 0$; and 2) closer to the jamming threshold ($\rho = 0.61$), winding of the plectonemes beyond $|\sigma| = 0.027$ even provokes significant decompaction of the DNA coil. This latter point is surprising because it contrasts with the monotonous behavior of $\langle d \rangle$ and $\langle Wr/\Delta Lk \rangle$ with respect to $|\sigma|$ and ρ that is observed in Fig. 4. Indeed, it was suggested above that a decrease in $\langle d \rangle$ and an increase in $\langle Wr/\Delta Lk \rangle$ facilitate compaction of the DNA coil. Then, why does the DNA coil expand for $\rho = 0.61$ and $|\sigma| > 0.027$, although $\langle d \rangle$ goes on decreasing and $\langle Wr/\Delta Lk \rangle$ goes on increasing? As anticipated in the Introduction, the reason is that increasing $|\sigma|$ not only reduces the diameter of plectonemes, which makes compaction of the DNA coil easier, but also increases their stiffness, which has the inverse effect of opposing compaction. The increase in the stiffness of plectonemes as a function of $|\sigma|$ was checked for short DNA chains with 200 beads (1500 bp) in the absence of any crowder. These chains are sufficiently short for branching of plectonemes not to occur and a single plectoneme to be observed at any time. At regular time intervals, the two extremities of the plectoneme were sought for, and the midcurve of the two opposite strands was computed. The directional correlation function of the segments of the midcurve was then averaged over many different conformations to get an estimate of the persistence length according to standard fitting procedures. The log of the directional correlation function is shown for different values of $|\sigma|$ in the main plot of Fig. S3, and the evolution of the fitted values of the persistence length ξ is shown as a function of $|\sigma|$ in the inset of the same figure. It is seen that the persistence length (and consequently, the formal bending rigidity) of the midcurve of the plectonemes increases by more than 50% from $|\sigma| = 0.035$ to $|\sigma| = 0.077$. All in all, Fig. 3 indicates that compaction of the DNA coil for $0.49 \leq \rho \leq 0.61$ and $|\sigma| > 0.027$ results from the balance of two conflicting trends, namely the decrease of the diameter of the plectonemes and the increase of their stiffness, with the increase in stiffness becoming predominant at larger crowder concentrations.

Departure from the spherical geometry at the jamming threshold

Finally, for topologically constrained DNA, a third regime is observed at crowder concentrations very close to the jamming threshold. Indeed, in all the simulations with $\sigma = 0$ or $\rho \leq 0.61$, the DNA chain relaxed toward a nearly spherical coil. This is illustrated in Fig. 5, which displays representative equilibrated conformations obtained with $\sigma = -0.078$ and $\rho = 0.61$ (first row) and $\sigma = 0$ and $\rho = 0.65$ (second row). In the left (respectively, right) vignettes, the DNA

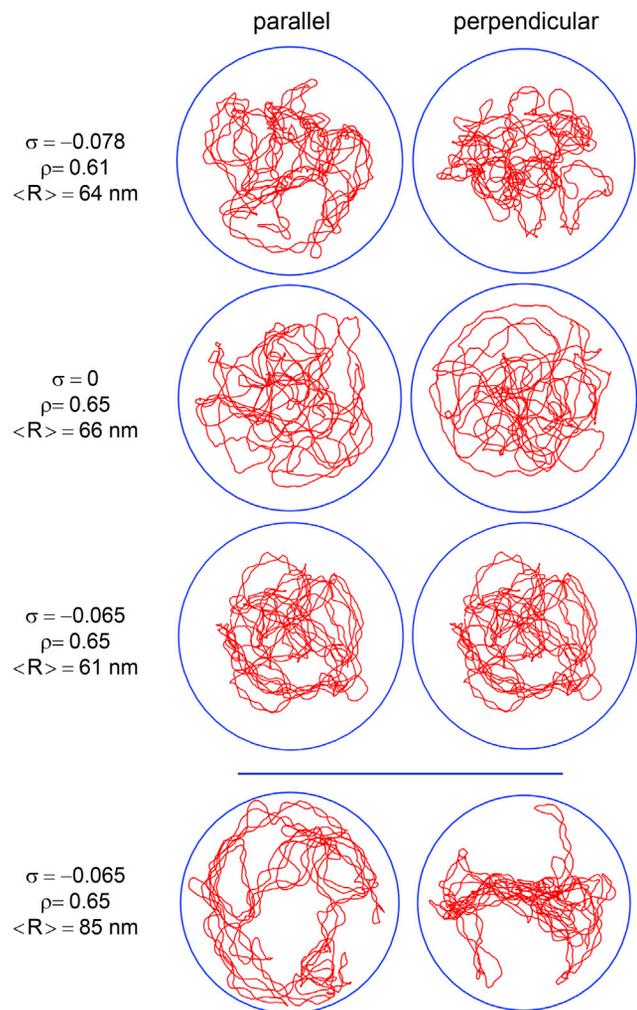


FIGURE 5 Representative snapshots viewed either parallel (*left column*) or perpendicular (*right column*) to the main axis of inertia of the DNA chain. Trajectories were run with $N = 1875$ ($\rho = 0.61$) or $N = 2000$ ($\rho = 0.65$) and different values of σ . The solid red line connects the centers of successive DNA beads. Crowding spheres are not shown. The blue circle shows the limits of the confinement sphere. Note the approximate spherical geometry of the DNA coil in the three top lines and its approximate toroidal geometry in the bottom line. To see this figure in color, go online.

coil is viewed parallel (respectively, *perpendicular*) to its principal axis of inertia with largest momentum. Because of the approximate spherical geometry, the aspect of the DNA coil does not change significantly from one viewpoint to the other one. Such nearly spherical DNA coils were also systematically observed in previous studies based on a somewhat different model of torsionally relaxed circular DNA (32,33). In contrast, for topologically constrained DNA ($|\sigma| \geq 0.027$) close to the jamming threshold ($\rho = 0.65$), we observed in this work that trajectories may relax either toward nearly spherical DNA coils or a fundamentally different type of nearly toroidal coils. This is illustrated in Fig. 5, in which the third and fourth rows display representative equilibrated conformations obtained with $\sigma = -0.065$ and $\rho = 0.65$. While in the third row the

geometry of the DNA coil is again nearly spherical, so that the coil has the same aspect when viewed from any axis, this is no longer the case in the fourth row, in which the geometry of the DNA coil is nearly toroidal, so that the coil appears like a ring when viewed parallel to the axis with largest momentum and like a disk when viewed perpendicular to this axis. As far as we can tell from 80 ms trajectories, both the spherical and toroidal conformations are stable or at least metastable. Because the toroidal geometry was never observed for torsionally relaxed DNA, it is most likely that the probability for the DNA chain to relax toward the toroidal geometry increases with $|\sigma|$. The rationale behind this observation is most probably that the toroidal geometry reduces the bending energy of the DNA coil by allowing the DNA chain to form large loops with radius close to that of the confinement sphere while still allowing demixing and compaction perpendicular to the plane of the torus. It is therefore expected that increasing $|\sigma|$, thereby making plectonemes more rigid, should favor relaxation toward the toroidal geometry. Quite importantly, the mean radius of equilibrated DNA coils with toroidal geometry is of the order of 85–90 nm, which is larger than the mean radius of torsionally relaxed DNA coils in the absence of any crowder (~ 82 nm; see Fig. 3). Still, such an increase in the mean radius of the DNA coil does not imply decompaction of the DNA coil because the DNA still occupies only a limited portion of the confinement sphere. Results obtained with $|\sigma| > 0$ and $\rho = 0.65$, at which the toroidal geometry is predominant, were therefore not displayed in Figs. 3 and 4 because $\langle R \rangle$ is no longer a measure of the compaction of the DNA coil. Last but not least, it may be worth mentioning that toroidal DNA coils are also obtained in the absence of crowders and confinement when explicit and relatively strong attraction between DNA segments is plugged explicitly into a model of torsionally relaxed DNA (see, for example, (4)). This is, of course, completely different from what is observed in the fourth row of Fig. 5, in which the toroidal geometry of the DNA coil is a direct consequence of the spherical geometry of the confinement chamber, which is specific to cocci. For nonspherical confinement chambers, such as the capped cylinders specific to bacilli (including *E. coli*), DNA coils will instead relax toward more complex geometries that minimize both the bending energy of the DNA chain and the volume occupied by the coil.

CONCLUSIONS

The demixing of DNA from other macromolecules of the cytosol and the formation of plectonemes are two independent mechanisms that are both able to compact the DNA coil. In this work, we investigated the interplay of these two mechanisms through coarse-grained modeling and Brownian dynamics simulations with the goal of understanding how a topologically constrained DNA molecule com-

pacts under the influence of noninteracting globular crowders. The model suggests that there exist three different regimes, depending on the superhelical density σ of the DNA molecule and the effective volume ratio ρ of the crowders:

- 1) Below a certain threshold for $|\sigma|$, the effects of the two mechanisms are additive, and the total compaction ratio of the DNA coil is the sum of the ratios due to demixing and supercoiling. Compaction of supercoiled DNA is facilitated by the decrease of the diameter of the plectonemes with increasing values of ρ .
- 2) Above this threshold for $|\sigma|$, the mean radius of the DNA coil ceases to decrease with increasing values of $|\sigma|$ because of the increasing stiffness of the plectonemes. For sufficiently large values of ρ , the DNA coil even decompacts upon increase of $|\sigma|$.
- 3) For values of ρ just below the jamming threshold, the coil formed by topologically constrained DNA may adopt a nonspherical geometry, which represents some trade-off between the minimization of the bending energy and the minimization of the volume of the coil. For example, toroidal DNA coils were observed with the spherical confinement chamber.

According to the model, the threshold where the two mechanisms cease to have additive effects on the compaction of the DNA coil lies around $|\sigma| = 0.027$. This turns out to be almost exactly the value of the effective supercoil density that is observed in living cells ($\sigma = -0.025$), where DNA-binding proteins reduce the number of supercoils to approximately one-half of the value in protein-free samples (90,91). Owing to the approximations of the model, such an exact correspondence is likely to be fortuitous, but it still suggests that both the additive and the nonadditive regime may be relevant in vivo because most biological functions rely on alternative winding and unwinding of the circular DNA (92,93). Moreover, the translational diffusion coefficient of macromolecules is much smaller in bacterial cells than in water and in eukaryotic cells (94), which indicates that the bacterial cytosol is indeed close to jamming. The regime predicted by the model, in which the mean radius of the nucleoid increases with $|\sigma|$ instead of decreasing, and the abrupt change of its geometry very close to the jamming threshold may consequently also be relevant for living cells. Finally, although the explicit modeling of topological domains is beyond the scope of this work, the results presented above support the hypothesis that differences in DNA concentration between different macrodomains (51–57) may indeed reflect differences in the level of superhelical density of the corresponding topological domains (58,59), provided that these domains are in the additive regime. An interesting related question is that of topological insulators and, more precisely, of the nature of the constraints these proteins must exert on the DNA duplexes to which they bind to prevent the diffusion of DNA supercoils (95–98). Work in this direction is in progress.

More generally, it may be worth emphasizing that there exist, in addition to DNA demixing and supercoiling, several other mechanisms that may contribute to the compaction of the bacterial DNA and the formation of the nucleoid (for a recent review, see (4)). Each of these mechanisms, which have not been taken into account in the model proposed here, may interact additively or destructively with DNA demixing and supercoiling. Of particular interest are the actions of nucleoid proteins, which can bridge (like H-NS), bend (like IHF, HU, and Fis), or wrap (like Dps) the DNA molecule (99). Cells lacking both HU and Fis have a large decondensed nucleoid (100), whereas the overproduction of H-NS leads instead to very compact nucleoids and may be lethal (101). Moreover, most nucleoid proteins such as Fis (102,103), HU (104–106), H-NS (102,107), and IHF (108) are capable of inducing gradual and strong DNA compaction *in vitro*, although at concentrations much larger than *in vivo* ones (109–111). It is known that nucleoid proteins interact with the underwinding of the DNA molecule (112) and are responsible for the fact that the number of supercoils in living cells is approximately one-half of the value in protein-free samples (90,91). It may therefore be interesting in future work to introduce DNA-binding proteins in the model—as was done, for example, in (82–84)—to investigate the interplay of DNA demixing and supercoiling with the binding of nucleoid proteins. In contrast, compaction of the DNA molecule in eukaryotic cells is primarily due to its wrapping around histone proteins, with supercoiling and crowding by nonbinding macromolecules playing a priori a minor role compared to prokaryotes. This problem is therefore rather different from the formation of the bacterial nucleoid, and its study requires the development of quite different models (see, for example, (113)).

SUPPORTING MATERIAL

Supporting Material can be found online at <https://doi.org/10.1016/j.bpj.2019.09.026>.

SUPPORTING CITATIONS

Reference (114) appears in the Supporting Material.

REFERENCES

- Zimmerman, S. B. 2006. Shape and compaction of *Escherichia coli* nucleoids. *J. Struct. Biol.* 156:255–261.
- de Vries, R. 2010. DNA condensation in bacteria: interplay between macromolecular crowding and nucleoid proteins. *Biochimie.* 92:1715–1721.
- Benza, V. G., B. Bassetti, ..., M. C. Lagomarsino. 2012. Physical descriptions of the bacterial nucleoid at large scales, and their biological implications. *Rep. Prog. Phys.* 75:076602.
- Joyeux, M. 2015. Compaction of bacterial genomic DNA: clarifying the concepts. *J. Phys. Condens. Matter.* 27:383001.
- Joyeux, M. 2016. *In vivo* compaction dynamics of bacterial DNA: a fingerprint of DNA/RNA demixing? *Curr. Opin. Colloid Interface Sci.* 26:17–27.
- Sherratt, D. J. 2003. Bacterial chromosome dynamics. *Science.* 301:780–785.
- Endesfelder, U., K. Finan, ..., M. Heilemann. 2013. Multiscale spatial organization of RNA polymerase in *Escherichia coli*. *Biophys. J.* 105:172–181.
- Hadizadeh Yazdi, N., C. C. Guet, ..., J. F. Marko. 2012. Variation of the folding and dynamics of the *Escherichia coli* chromosome with growth conditions. *Mol. Microbiol.* 86:1318–1333.
- Jin, D. J., C. Cagliero, ..., Y. N. Zhou. 2015. The dynamic nature and territory of transcriptional machinery in the bacterial chromosome. *Front. Microbiol.* 6:497.
- Jin, D. J., C. Cagliero, and Y. N. Zhou. 2013. Role of RNA polymerase and transcription in the organization of the bacterial nucleoid. *Chem. Rev.* 113:8662–8682.
- Stracy, M., C. Lesterlin, ..., A. N. Kapanidis. 2015. Live-cell super-resolution microscopy reveals the organization of RNA polymerase in the bacterial nucleoid. *Proc. Natl. Acad. Sci. USA.* 112:E4390–E4399.
- Spahn, C., U. Endesfelder, and M. Heilemann. 2014. Super-resolution imaging of *Escherichia coli* nucleoids reveals highly structured and asymmetric segregation during fast growth. *J. Struct. Biol.* 185:243–249.
- Fisher, J. K., A. Bourniquel, ..., N. Kleckner. 2013. Four-dimensional imaging of *E. coli* nucleoid organization and dynamics in living cells. *Cell.* 153:882–895.
- Cagliero, C. and D. J. Jin. 2013. Dissociation and re-association of RNA polymerase with DNA during osmotic stress response in *Escherichia coli*. *Nucleic Acids Res.* 41:315–326.
- Cabrera, J. E., C. Cagliero, ..., D. J. Jin. 2009. Active transcription of rRNA operons condenses the nucleoid in *Escherichia coli*: examining the effect of transcription on nucleoid structure in the absence of transcription. *J. Bacteriol.* 191:4180–4185.
- Bakshi, S., H. Choi, and J. C. Weisshaar. 2015. The spatial biology of transcription and translation in rapidly growing *Escherichia coli*. *Front. Microbiol.* 6:636.
- Bakshi, S., A. Siryaporn, ..., J. C. Weisshaar. 2012. Superresolution imaging of ribosomes and RNA polymerase in live *Escherichia coli* cells. *Mol. Microbiol.* 85:21–38.
- Bakshi, S., H. Choi, ..., J. C. Weisshaar. 2014. Time-dependent effects of transcription- and translation-halting drugs on the spatial distributions of the *Escherichia coli* chromosome and ribosomes. *Mol. Microbiol.* 94:871–887.
- Teraoka, I. 2002. *Polymer Solutions: An Introduction to Physical Properties.* Wiley, New-York.
- Sear, R. P. 1998. Coil-globule transition of a semiflexible polymer driven by the addition of spherical particles. *Phys. Rev. E Stat. Phys. Plasmas Fluids Relat. Interdiscip. Topics.* 58:724–728.
- van der Schoot, P. 1998. Protein-induced collapse of polymer chains. *Macromolecules.* 31:4635–4638.
- Odijk, T. 1998. Osmotic compaction of supercoiled DNA into a bacterial nucleoid. *Biophys. Chem.* 73:23–29.
- Castelnovo, M., and W. M. Gelbart. 2004. Semiflexible chain condensation by neutral depleting agents: role of correlations between depletants. *Macromolecules.* 37:3510–3517.
- de Vries, R. 2006. Depletion-induced instability in protein-DNA mixtures: influence of protein charge and size. *J. Chem. Phys.* 125:014905.
- Krotova, M. K., V. V. Vasilevskaya, ..., A. R. Khokhlov. 2010. DNA compaction in a crowded environment with negatively charged proteins. *Phys. Rev. Lett.* 105:128302.
- Yoshikawa, K., S. Hirota, ..., Y. Yoshikawa. 2010. Compaction of DNA induced by like-charge protein: opposite salt-effect against the polymer-salt-induced condensation with neutral polymer. *J. Phys. Chem. Lett.* 1:1763–1766.

27. Zinchenko, A., K. Tsumoto, ..., K. Yoshikawa. 2014. Crowding by anionic nanoparticles causes DNA double-strand instability and compaction. *J. Phys. Chem. B*. 118:1256–1262.
28. Kim, J., C. Jeon, ..., B. Y. Ha. 2015. A polymer in a crowded and confined space: effects of crowder size and poly-dispersity. *Soft Matter*. 11:1877–1888.
29. Jeon, C., Y. Jung, and B. Y. Ha. 2016. Effects of molecular crowding and confinement on the spatial organization of a biopolymer. *Soft Matter*. 12:9436–9450.
30. Jeon, C., C. Hyeon, ..., B. Y. Ha. 2016. How are molecular crowding and the spatial organization of a biopolymer interrelated. *Soft Matter*. 12:9786–9796.
31. Jeon, C., Y. Jung, and B. Y. Ha. 2017. A ring-polymer model shows how macromolecular crowding controls chromosome-arm organization in *Escherichia coli*. *Sci. Rep.* 7:11896.
32. Joyeux, M. 2017. Coarse-grained model of the demixing of DNA and non-binding globular macromolecules. *J. Phys. Chem. B*. 121:6351–6358.
33. Joyeux, M. 2018. A segregative phase separation scenario of the formation of the bacterial nucleoid. *Soft Matter*. 14:7368–7381.
34. Hsu, C. C., and J. M. Prausnitz. 1974. Thermodynamics of polymer compatibility in ternary systems. *Macromolecules*. 7:320–324.
35. Mutch, K. J., J. S. van Duijneveldt, and J. Eastoe. 2007. Colloid–polymer mixtures in the protein limit. *Soft Matter*. 3:155–167.
36. Sear, R. P. 2001. Phase separation in mixtures of colloids and long ideal polymer coils. *Phys. Rev. Lett.* 86:4696–4699.
37. Odijk, T. 1997. Many-body depletion interactions among protein spheres in a semidilute polymer solution. *J. Chem. Phys.* 106:3402–3406.
38. Fleer, G. J., and R. Tuinier. 2007. Analytical phase diagram for colloid-polymer mixtures. *Phys. Rev. E Stat. Nonlin. Soft Matter Phys.* 76:041802.
39. Bolhuis, P. G., E. J. Meijer, and A. A. Louis. 2003. Colloid-polymer mixtures in the protein limit. *Phys. Rev. Lett.* 90:068304.
40. Fuchs, M., and K. S. Schweizer. 2002. Structure of colloid–polymer suspensions. *J. Phys. Condens. Matter*. 14:R239–R269.
41. Fuchs, M., and K. S. Schweizer. 2000. Structure and thermodynamics of colloid-polymer mixtures: a macromolecular approach. *Europhys. Lett.* 51:621–627.
42. Fuchs, M., and K. S. Schweizer. 2001. Macromolecular theory of solvation and structure in mixtures of colloids and polymers. *Phys. Rev. E Stat. Nonlin. Soft Matter Phys.* 64:021514.
43. Chen, Y.-L., and K. S. Schweizer. 2002. Depletion interactions in suspensions of spheres and rod-polymers. *J. Chem. Phys.* 117:1351–1362.
44. Kulkarni, A. M., A. P. Chatterjee, ..., C. F. Zukowski. 1999. Depletion interactions in the protein limit: effects of polymer density fluctuations. *Phys. Rev. Lett.* 83:4554–4557.
45. Woodward, C. E., and J. Forsman. 2015. A many-body Hamiltonian for nanoparticles immersed in a polymer solution. *Langmuir*. 31:22–26.
46. Bates, A. D., and A. Maxwell. 2005. DNA Topology. Oxford University Press, New York.
47. Stuger, R., C. L. Woldringh, ..., H. V. Westerhoff. 2002. DNA supercoiling by gyrase is linked to nucleoid compaction. *Mol. Biol. Rep.* 29:79–82.
48. Cunha, S., C. L. Woldringh, and T. Odijk. 2001. Polymer-mediated compaction and internal dynamics of isolated *Escherichia coli* nucleoids. *J. Struct. Biol.* 136:53–66.
49. Boles, T. C., J. H. White, and N. R. Cozzarelli. 1990. Structure of plectonemically supercoiled DNA. *J. Mol. Biol.* 213:931–951.
50. Bednar, J., P. Furrer, ..., A. D. Bates. 1994. The twist, writhe and overall shape of supercoiled DNA change during counterion-induced transition from a loosely to a tightly interwound superhelix. Possible implications for DNA structure in vivo. *J. Mol. Biol.* 235:825–847.
51. Niki, H., Y. Yamaichi, and S. Hiraga. 2000. Dynamic organization of chromosomal DNA in *Escherichia coli*. *Genes Dev.* 14:212–223.
52. Valens, M., S. Penaud, ..., F. Boccard. 2004. Macrodome organization of the *Escherichia coli* chromosome. *EMBO J.* 23:4330–4341.
53. Espéli, O., R. Mercier, and F. Boccard. 2008. DNA dynamics vary according to macrodomain topography in the *E. coli* chromosome. *Mol. Microbiol.* 68:1418–1427.
54. Duigou, S., and F. Boccard. 2017. Long range chromosome organization in *Escherichia coli*: the position of the replication origin defines the non-structured regions and the right and left macrodomains. *PLoS Genet.* 13:e1006758.
55. Mercier, R., M. A. Petit, ..., O. Espéli. 2008. The MatP/matS site-specific system organizes the terminus region of the *E. coli* chromosome into a macrodomain. *Cell*. 135:475–485.
56. Dupaigne, P., N. K. Tonthat, ..., M. A. Schumacher. 2012. Molecular basis for a protein-mediated DNA-bridging mechanism that functions in condensation of the *E. coli* chromosome. *Mol. Cell*. 48:560–571.
57. Valens, M., A. Thiel, and F. Boccard. 2016. The MaoP/maoS site-specific system organizes the Ori region of the *E. coli* chromosome into a macrodomain. *PLoS Genet.* 12:e1006309.
58. Postow, L., C. D. Hardy, ..., N. R. Cozzarelli. 2004. Topological domain structure of the *Escherichia coli* chromosome. *Genes Dev.* 18:1766–1779.
59. Deng, S., R. A. Stein, and N. P. Higgins. 2005. Organization of supercoil domains and their reorganization by transcription. *Mol. Microbiol.* 57:1511–1521.
60. Lyubchenko, Y. L., and L. S. Shlyakhtenko. 1997. Visualization of supercoiled DNA with atomic force microscopy in situ. *Proc. Natl. Acad. Sci. USA*. 94:496–501.
61. Hammermann, M., N. Brun, ..., J. Langowski. 1998. Salt-dependent DNA superhelix diameter studied by small angle neutron scattering measurements and Monte Carlo simulations. *Biophys. J.* 75:3057–3063.
62. Zakharova, S. S., W. Jesse, ..., J. R. van der Maarel. 2002. Dimensions of plectonemically supercoiled DNA. *Biophys. J.* 83:1106–1118.
63. Latulippe, D. R., and A. L. Zydney. 2010. Radius of gyration of plasmid DNA isoforms from static light scattering. *Biotechnol. Bioeng.* 107:134–142.
64. Irobalieva, R. N., J. M. Fogg, ..., L. Zechiedrich. 2015. Structural diversity of supercoiled DNA. *Nat. Commun.* 6:8440.
65. Marko, J. F., and E. D. Siggia. 1995. Statistical mechanics of supercoiled DNA. *Phys. Rev. E Stat. Phys. Plasmas Fluids Relat. Interdiscip. Topics*. 52:2912–2938.
66. Vologodskii, A. V., and N. R. Cozzarelli. 1994. Conformational and thermodynamic properties of supercoiled DNA. *Annu. Rev. Biophys. Biomol. Struct.* 23:609–643.
67. Vologodskii, A., and N. R. Cozzarelli. 1996. Effect of supercoiling on the juxtaposition and relative orientation of DNA sites. *Biophys. J.* 70:2548–2556.
68. Podtelezhnikov, A. A., N. R. Cozzarelli, and A. V. Vologodskii. 1999. Equilibrium distributions of topological states in circular DNA: interplay of supercoiling and knotting. *Proc. Natl. Acad. Sci. USA*. 96:12974–12979.
69. Krajina, B. A., and A. J. Spakowitz. 2016. Large-scale conformational transitions in supercoiled DNA revealed by coarse-grained simulation. *Biophys. J.* 111:1339–1349.
70. Fathizadeh, A., H. Schiessel, and M. R. Ejtehadi. 2015. Molecular dynamics simulation of supercoiled DNA rings. *Macromolecules*. 48:164–172.
71. Schlick, T., and W. K. Olson. 1992. Supercoiled DNA energetics and dynamics by computer simulation. *J. Mol. Biol.* 223:1089–1119.
72. Benedetti, F., A. Japaridze, ..., A. Stasiak. 2015. Effects of physiological self-crowding of DNA on shape and biological properties of DNA molecules with various levels of supercoiling. *Nucleic Acids Res.* 43:2390–2399.

73. Lim, W., S. Y. Ng, ..., J. R. van der Maarel. 2008. Conformational response of supercoiled DNA to confinement in a nanochannel. *J. Chem. Phys.* 129:165102.
74. Sottas, P. E., E. Larquet, ..., J. Dubochet. 1999. Brownian dynamics simulation of DNA condensation. *Biophys. J.* 77:1858–1870.
75. Marko, J. F. 2011. Scaling of linking and writhing numbers for spherically confined and topologically equilibrated flexible polymers. *J. Stat. Phys.* 142:1353–1370.
76. Zhu, X., S. Y. Ng, ..., J. R. van der Maarel. 2010. Effect of crowding on the conformation of interwound DNA strands from neutron scattering measurements and Monte Carlo simulations. *Phys. Rev. E Stat. Nonlin. Soft Matter Phys.* 81:061905.
77. Gupta, A. N., and J. R. van der Maarel. 2017. Compaction of plasmid DNA by macromolecular crowding. *Macromolecules.* 50:1666–1671.
78. Ramos, J. E., Jr., J. R. Neto, and R. de Vries. 2008. Polymer induced condensation of DNA supercoils. *J. Chem. Phys.* 129:185102.
79. Florescu, A. M., and M. Joyeux. 2009. Description of nonspecific DNA-protein interaction and facilitated diffusion with a dynamical model. *J. Chem. Phys.* 130:015103.
80. Florescu, A. M., and M. Joyeux. 2009. Dynamical model of DNA protein interaction: effect of protein charge distribution and mechanical properties. *J. Chem. Phys.* 131:105102.
81. Florescu, A. M., and M. Joyeux. 2010. Comparison of kinetic and dynamical models of DNA-protein interaction and facilitated diffusion. *J. Phys. Chem. A.* 114:9662–9672.
82. Joyeux, M., and J. Vreede. 2013. A model of H-NS mediated compaction of bacterial DNA. *Biophys. J.* 104:1615–1622.
83. Joyeux, M. 2014. Equilibration of complexes of DNA and H-NS proteins on charged surfaces: a coarse-grained model point of view. *J. Chem. Phys.* 141:115102.
84. Joyeux, M. 2018. Role of salt valency in the switch of H-NS proteins between DNA-bridging and DNA-stiffening modes. *Biophys. J.* 114:2317–2325.
85. Joyeux, M. 2019. Preferential localization of the bacterial nucleoid. *Microorganisms.* 7:204.
86. Chirico, G., and J. Langowski. 1994. Kinetics of DNA supercoiling studied by Brownian dynamics simulation. *Biopolymers.* 34:415–433.
87. Bauer, W. R. 1978. Structure and reactions of closed duplex DNA. *Annu. Rev. Biophys. Bioeng.* 7:287–313.
88. Manning, G. S. 1969. Limiting laws and counterion condensation in polyelectrolyte solutions. I. Colligative properties. *J. Chem. Phys.* 51:924–933.
89. Oosawa, F. 1971. Polyelectrolytes. Marcel Dekker, New York.
90. Bliska, J. B., and N. R. Cozzarelli. 1987. Use of site-specific recombination as a probe of DNA structure and metabolism in vivo. *J. Mol. Biol.* 194:205–218.
91. Zacharias, W., A. Jaworski, ..., R. D. Wells. 1988. The B- to Z-DNA equilibrium in vivo is perturbed by biological processes. *Proc. Natl. Acad. Sci. USA.* 85:7069–7073.
92. Travers, A., and G. Muskhelishvili. 2005. DNA supercoiling - a global transcriptional regulator for enterobacterial growth? *Nat. Rev. Microbiol.* 3:157–169.
93. Hatfield, G. W., and C. J. Benham. 2002. DNA topology-mediated control of global gene expression in *Escherichia coli*. *Annu. Rev. Genet.* 36:175–203.
94. Mika, J. T., and B. Poolman. 2011. Macromolecule diffusion and confinement in prokaryotic cells. *Curr. Opin. Biotechnol.* 22:117–126.
95. Moulin, L., A. R. Rahmouni, and F. Boccard. 2005. Topological insulators inhibit diffusion of transcription-induced positive supercoils in the chromosome of *Escherichia coli*. *Mol. Microbiol.* 55:601–610.
96. Leng, F., B. Chen, and D. D. Dunlap. 2011. Dividing a supercoiled DNA molecule into two independent topological domains. *Proc. Natl. Acad. Sci. USA.* 108:19973–19978.
97. Yan, Y., Y. Ding, ..., L. Finzi. 2018. Protein-mediated loops in supercoiled DNA create large topological domains. *Nucleic Acids Res.* 46:4417–4424.
98. Xia, Y., K. W. Zheng, ..., Z. Tan. 2018. Transmission of dynamic supercoiling in linear and multi-way branched DNAs and its regulation revealed by a fluorescent G-quadruplex torsion sensor. *Nucleic Acids Res.* 46:7418–7424.
99. Luijsterburg, M. S., M. F. White, ..., R. T. Dame. 2008. The major architects of chromatin: architectural proteins in bacteria, archaea and eukaryotes. *Crit. Rev. Biochem. Mol. Biol.* 43:393–418.
100. Paull, T. T., and R. C. Johnson. 1995. DNA looping by *Saccharomyces cerevisiae* high mobility group proteins NHP6A/B. Consequences for nucleoprotein complex assembly and chromatin condensation. *J. Biol. Chem.* 270:8744–8754.
101. Spurio, R., M. Dürrenberger, ..., C. O. Gualerzi. 1992. Lethal overproduction of the *Escherichia coli* nucleoid protein H-NS: ultramicroscopic and molecular autopsy. *Mol. Gen. Genet.* 231:201–211.
102. Schneider, R., R. Lurz, ..., G. Muskhelishvili. 2001. An architectural role of the *Escherichia coli* chromatin protein FIS in organising DNA. *Nucleic Acids Res.* 29:5107–5114.
103. Skoko, D., D. Yoo, ..., R. C. Johnson. 2006. Mechanism of chromosome compaction and looping by the *Escherichia coli* nucleoid protein Fis. *J. Mol. Biol.* 364:777–798.
104. Zentgraf, H., V. Berthold, and K. Geider. 1977. Interaction of DNA with DNA binding proteins. II. Displacement of *Escherichia coli* DNA unwinding protein and the condensed structure of DNA complexed with protein HD. *Biochim. Biophys. Acta.* 474:629–638.
105. Endo, T., N. Sasaki, ..., M. Nakata. 2002. Compact form of DNA induced by DNA-binding protein HU. *Biochem. Biophys. Res. Commun.* 290:546–551.
106. Sagi, D., N. Friedman, ..., J. Stavans. 2004. Modulation of DNA conformations through the formation of alternative high-order HU-DNA complexes. *J. Mol. Biol.* 341:419–428.
107. Dame, R. T., C. Wyman, and N. Goosen. 2000. H-NS mediated compaction of DNA visualised by atomic force microscopy. *Nucleic Acids Res.* 28:3504–3510.
108. Lin, J., H. Chen, ..., J. Yan. 2012. Physical organization of DNA by multiple non-specific DNA-binding modes of integration host factor (IHF). *PLoS One.* 7:e49885.
109. Pettijohn, D. E. 1990. Bacterial chromosome structure. *Nucleic Acids Mol. Biol.* 4:152–162.
110. Rouvière-Yaniv, J., M. Yaniv, and J. E. Germond. 1979. *E. coli* DNA binding protein HU forms nucleosomal-like structure with circular double-stranded DNA. *Cell.* 17:265–274.
111. Geider, K., and H. Hoffmann-Berling. 1981. Proteins controlling the helical structure of DNA. *Annu. Rev. Biochem.* 50:233–260.
112. Lim, C. J., L. J. Kenney, and J. Yan. 2014. Single-molecule studies on the mechanical interplay between DNA supercoiling and H-NS DNA architectural properties. *Nucleic Acids Res.* 42:8369–8378.
113. Bascom, G. D., and T. Schlick. 2018. Mesoscale modeling of chromatin fibers. In *Nuclear Architecture and Dynamics*. C. Lavelle and J.-M. Victor, eds. Academic Press, pp. 123–147.
114. Jian, H., A. Vologodskii, and T. Schlick. 1997. A combined wormlike-chain and bead model for dynamic simulations of long linear DNA. *J. Comput. Phys.* 136:168–179.

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Supplemental Information

Bacterial Nucleoid: Interplay of DNA Demixing and Supercoiling

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Bacterial nucleoid: Interplay of DNA demixing and supercoiling

- Supporting Material -

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MODEL AND SIMULATIONS

Temperature T is assumed to be 298 K throughout the study. The model consists of a circular chain of $n = 2880$ beads with radius $a = 1.0$ nm separated at equilibrium by a distance $l_0 = 2.5$ nm (the genomic DNA) enclosed in a large confinement sphere of radius $R_0 = 120$ nm (the cell), together with N spheres of radius $b = 7.4$ nm (the crowding globular macromolecules). Two beads represent 15 DNA base pairs. The contour length of the DNA chain and the volume of the confinement sphere correspond approximately to 1/200th of the values for *E. coli* cells, so that the DNA concentration of the model is close to the physiological value (about 10 mM). The potential energy of the system, E_{pot} , consists of four terms

$$E_{\text{pot}} = V_{\text{DNA}} + V_{\text{DNA/C}} + V_{\text{C/C}} + V_{\text{wall}} , \quad (\text{S1})$$

which describe the internal energy of the DNA molecule, DNA-crowder interactions, crowder-crowder interactions, and the repulsive potential that maintains DNA beads and crowding spheres inside the confinement sphere, respectively. The internal energy of the DNA chain is further written as the sum of 4 contributions

$$V_{\text{DNA}} = \frac{\hbar}{2} \sum_{k=1}^n (l_k - l_0)^2 + \frac{g}{2} \sum_{k=1}^n \theta_k^2 + \frac{\tau}{2} \sum_{k=1}^n (\Phi_{k+1} - \Phi_k)^2 + q^2 \sum_{k=1}^{n-2} \sum_{K=k+2}^n H(\|\mathbf{r}_k - \mathbf{r}_K\| - 2a) , \quad (\text{S2})$$

where

$$H(r) = \frac{1}{4\pi\epsilon r} \exp\left(-\frac{r}{r_D}\right) , \quad (\text{S3})$$

which describe the stretching, bending, torsional, and electrostatic energy of the DNA chain, respectively. \mathbf{r}_k denotes the position of DNA bead k , $l_k = \|\mathbf{r}_k - \mathbf{r}_{k+1}\|$ the distance between two successive beads, and $\theta_k = \arccos((\mathbf{r}_k - \mathbf{r}_{k+1})(\mathbf{r}_{k+1} - \mathbf{r}_{k+2}) / (\|\mathbf{r}_k - \mathbf{r}_{k+1}\| \|\mathbf{r}_{k+1} - \mathbf{r}_{k+2}\|))$ the angle formed by three successive beads. The stretching energy is a computational device without biological meaning, which is aimed at avoiding a rigid rod description. The stretching force constant h was set to $h = 100 k_B T / l_0^2$ to insure that the variations of the distance between successive beads remain small enough (1). In contrast, the bending rigidity was obtained from the known persistence length of DNA, $\xi = 50$ nm, according to $g = \xi k_B T / l_0 = 20 k_B T$. The torsion contribution is borrowed from Ref. (2) and torsional forces and momenta are computed as described therein. $\Phi_{k+1} - \Phi_k$ denotes the rotation of the body-fixed frame $(\mathbf{u}_k, \mathbf{f}_k, \mathbf{v}_k)$ between DNA beads k and $k+1$. The value of the torsional rigidity, $\tau = 25 k_B T$, was obtained by imposing that the writhe contribution Wr accounts for approximately 70% of the linking number difference ΔLk at equilibrium (3), see Fig. 4. The value of the torsional rigidity used in the simulations is close to the value of the bending rigidity, which agrees with experimental findings (3). Finally, the electrostatic energy of the DNA chain is written as a sum of repulsive Debye-Hückel terms with hard core. $\epsilon = 80 \epsilon_0$ denotes the dielectric constant of the buffer and $r_D = 1.07$ nm the Debye length inside the buffer. This value of the Debye length corresponds to a concentration of monovalent salt of 100 mM, which is the value that is generally assumed for the cytoplasm of bacterial cells. q is the value of the electric charge, which is placed at the centre of each DNA bead

$$q = -\frac{l_0 \bar{e}}{\ell_B} \approx -3.5 \bar{e}, \quad (\text{S4})$$

where \bar{e} is the absolute charge of the electron and $\ell_B = 0.7$ nm the Bjerrum length of water. In Eq. (S4), \bar{e} / ℓ_B is the net linear charge density along a DNA molecule immersed in a buffer with monovalent cations derived from Manning's counterion condensation theory (4,5). Note that electrostatic interactions between nearest neighbours are not included in Eq. (S2) because it is considered that they are already accounted for in the stretching and bending terms.

DNA-crowder and crowder-crowder interactions are similarly expressed as sums of Debye-Hückel potentials with hard cores

$$V_{\text{DNA/C}} = q^2 \sum_{k=1}^n \sum_{j=1}^N H(\|\mathbf{r}_k - \mathbf{R}_j\| - a - b) \quad (\text{S5})$$

$$V_{\text{C/C}} = q^2 \sum_{j=1}^{N-1} \sum_{J=j+1}^N H(\|\mathbf{R}_j - \mathbf{R}_J\| - 2b),$$

where \mathbf{R}_j denotes the position of crowding sphere j , and a charge q is placed at the centre of each crowding sphere, as for DNA beads. The repulsion potential between a DNA bead and a crowding sphere is therefore the median of the repulsion potential between two DNA beads and the repulsion potential between two crowding spheres. According to previous work (6,7), strong compaction of torsionally relaxed DNA chains is consequently expected for effective volume fractions of the crowders

$$\rho = \frac{N(b + \Delta b)^3}{R_0^3}, \quad (\text{S6})$$

in the range $0.60 \leq \rho \leq 0.70$, that is close to the jamming threshold for hard spheres. In Eq. (S6), $b + \Delta b$ represents the effective radius of the crowding spheres, that is half the distance between the centres of two spheres at which the electrostatic repulsion energy is equal to the thermal energy $k_B T$. Numerical values of the parameters reported above lead to $\Delta b = 0.865$ nm, so that $\rho \approx 0.49, 0.57, 0.61$, and 0.65 for $N = 1500, 1750, 1875$, and 2000 , respectively.

Finally, V_{wall} is written in the form

$$V_{\text{wall}} = \zeta \left(\sum_{k=1}^n f(\|\mathbf{r}_k\|) + \sum_{j=1}^N f(\|\mathbf{R}_j\|) \right), \quad (\text{S7})$$

where the repulsive force constant ζ is set to $1000 k_B T$ and the function $f(r)$ is defined according to

$$\text{if } r \leq R_0 : f(r) = 0$$

$$\text{if } r > R_0 : f(r) = \left(\frac{r}{R_0} \right)^6 - 1. \quad (\text{S8})$$

The dynamics of the system was investigated by integrating numerically overdamped Langevin equations. Practically, the updated positions and torsion angles at time step $i+1$ are computed from the positions and torsion angles at time step i according to

$$\begin{aligned}
\mathbf{r}_k^{(i+1)} &= \mathbf{r}_k^{(i)} + \frac{\Delta t}{6\pi\eta a} \mathbf{f}_k^{(i)} + \sqrt{\frac{2 k_B T \Delta t}{6\pi\eta a}} x_k^{(i)} \\
\mathbf{R}_j^{(i+1)} &= \mathbf{R}_j^{(i)} + \frac{\Delta t}{6\pi\eta b} \mathbf{F}_j^{(i)} + \sqrt{\frac{2 k_B T \Delta t}{6\pi\eta b}} X_j^{(i)} \\
\Phi_k^{(i+1)} &= \Phi_k^{(i)} + \frac{\tau \Delta t}{4\pi\eta a^2 l_0} (\Phi_{k+1}^{(i)} - 2\Phi_k^{(i)} + \Phi_{k-1}^{(i)}),
\end{aligned} \tag{S9}$$

where $\mathbf{f}_k^{(i)}$ and $\mathbf{F}_j^{(i)}$ are vectors of inter-particle forces arising from the potential energy E_{pot} , $T = 298$ K is the temperature of the system, $x_k^{(i)}$ and $X_j^{(i)}$ are vectors of random numbers extracted from a Gaussian distribution of mean 0 and variance 1, $\eta = 0.00089$ Pa s is the viscosity of the buffer at 298 K, and $\Delta t = 10$ ps is the integration time step. After each integration step, the position of the centre of the sphere was slightly adjusted so as to coincide with the centre of mass of the DNA molecule. Trajectories were integrated for 10 ms for $N = 0$, 20 ms for $N = 1500$ and $N = 1750$, 30 ms for $N = 1875$, and up to 80 ms for $N = 2000$.

The twist difference ΔTw , the writhe Wr , and the linking number difference ΔLk , of the DNA chain were computed at regular time intervals according to (2)

$$\begin{aligned}
\Delta Tw &= \frac{1}{2\pi} \sum_{k=1}^n (\Phi_{k+1} - \Phi_k) \\
Wr &= \frac{1}{4\pi} \sum_{k=1}^n \sum_{K \neq k} \frac{[(\mathbf{r}_{k+1} - \mathbf{r}_k) \times (\mathbf{r}_{K+1} - \mathbf{r}_K)] \cdot (\mathbf{r}_k - \mathbf{r}_K)}{|\mathbf{r}_k - \mathbf{r}_K|^3} \\
\Delta Lk &= \Delta Tw + Wr,
\end{aligned} \tag{S10}$$

The superhelical density σ was subsequently estimated from

$$\sigma = \frac{\Delta Lk}{Lk_0}, \tag{S11}$$

where the linking number $Lk_0 = 7.5n/10.5$ is the ratio of the formal number of base pairs of the DNA chain and the mean number of base pairs per turn of the torsionally relaxed double helix. $Lk_0 \approx 2057$ for $n = 2880$ and $Lk_0 \approx 143$ for $n = 200$.

SUPPORTING REFERENCES

1. Jian, H., A. Vologodskii, and T. Schlick. 1997. A combined wormlike-chain and bead model for dynamic simulations of long linear DNA. *J. Comp. Phys.* 136:168-179.
2. Chirico, G., and J. Langowski. 1994. Kinetics of DNA supercoiling studied by Brownian dynamics simulation. *Biopolymers.* 34:415-433.
3. Boles, T. C., J. H. White, and N. R. Cozzarelli. 1990. Structure of plectonemically supercoiled DNA. *J. Mol. Biol.* 213:931-951.
4. Manning, G. S. 1969. Limiting laws and counterion condensation in polyelectrolyte solutions. I. Colligative properties. *J. Chem. Phys.* 51:924-933.
5. Oosawa, F. 1971. Polyelectrolytes. Marcel Dekker, New York.
6. Joyeux, M. 2017. Coarse-grained model of the demixing of DNA and non-binding globular macromolecules. *J. Phys. Chem. B.* 121:6351-6358.
7. Joyeux, M. 2018. A segregative phase separation scenario of the formation of the bacterial nucleoid. *Soft Matter.* 14:7368-7381.

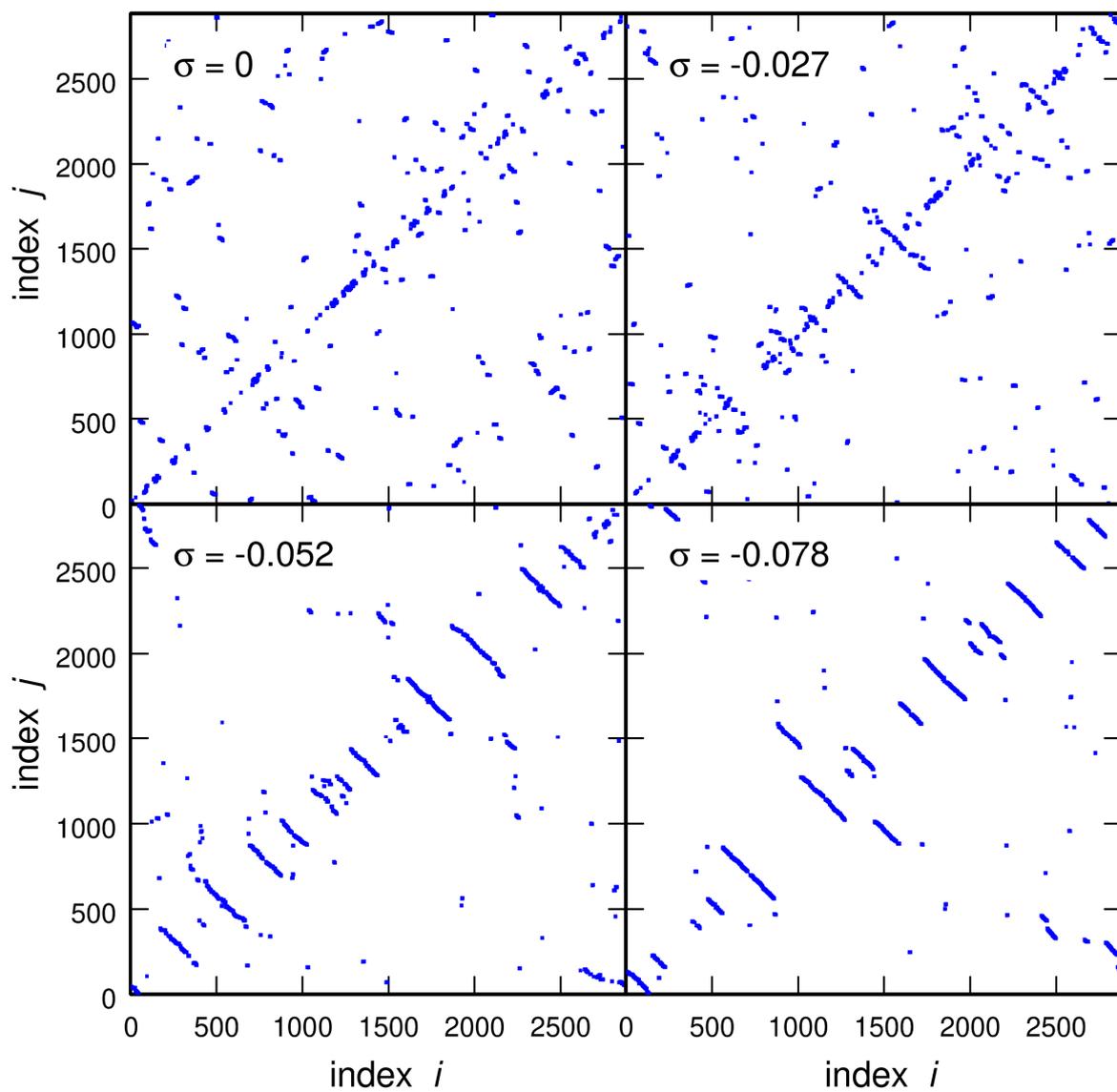


Figure S1 : Plot of the index j of the bead located closest to bead i for the equilibrated DNA conformations shown in Fig. 1. The segments of points parallel to the anti-diagonal denote the presence of plectonemes.

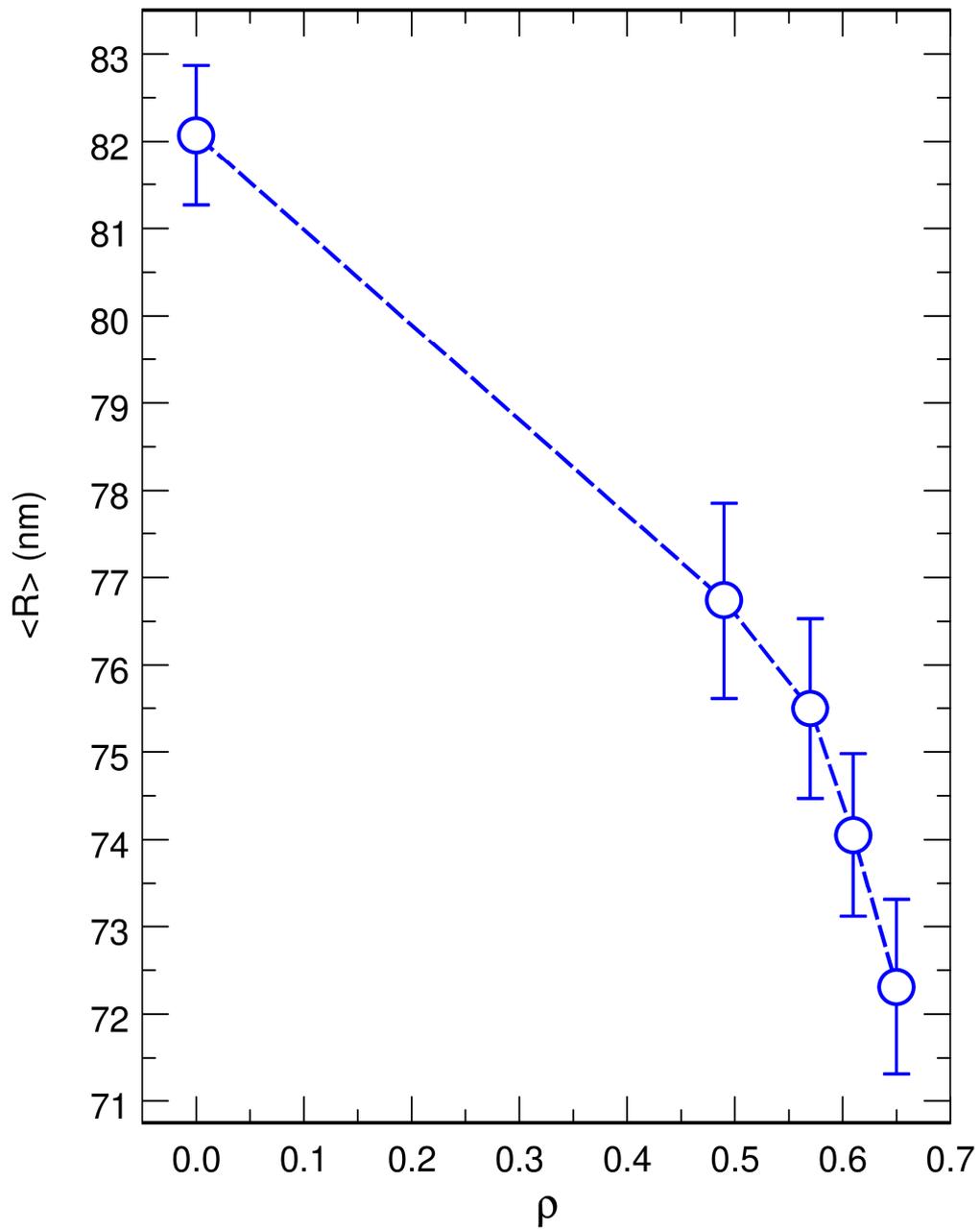


Figure S2 : Plot, as a function of the effective crowder volume ratio ρ , of the mean radius of the DNA coil $\langle R \rangle$ for torsionally relaxed DNA chains ($\sigma = 0$).

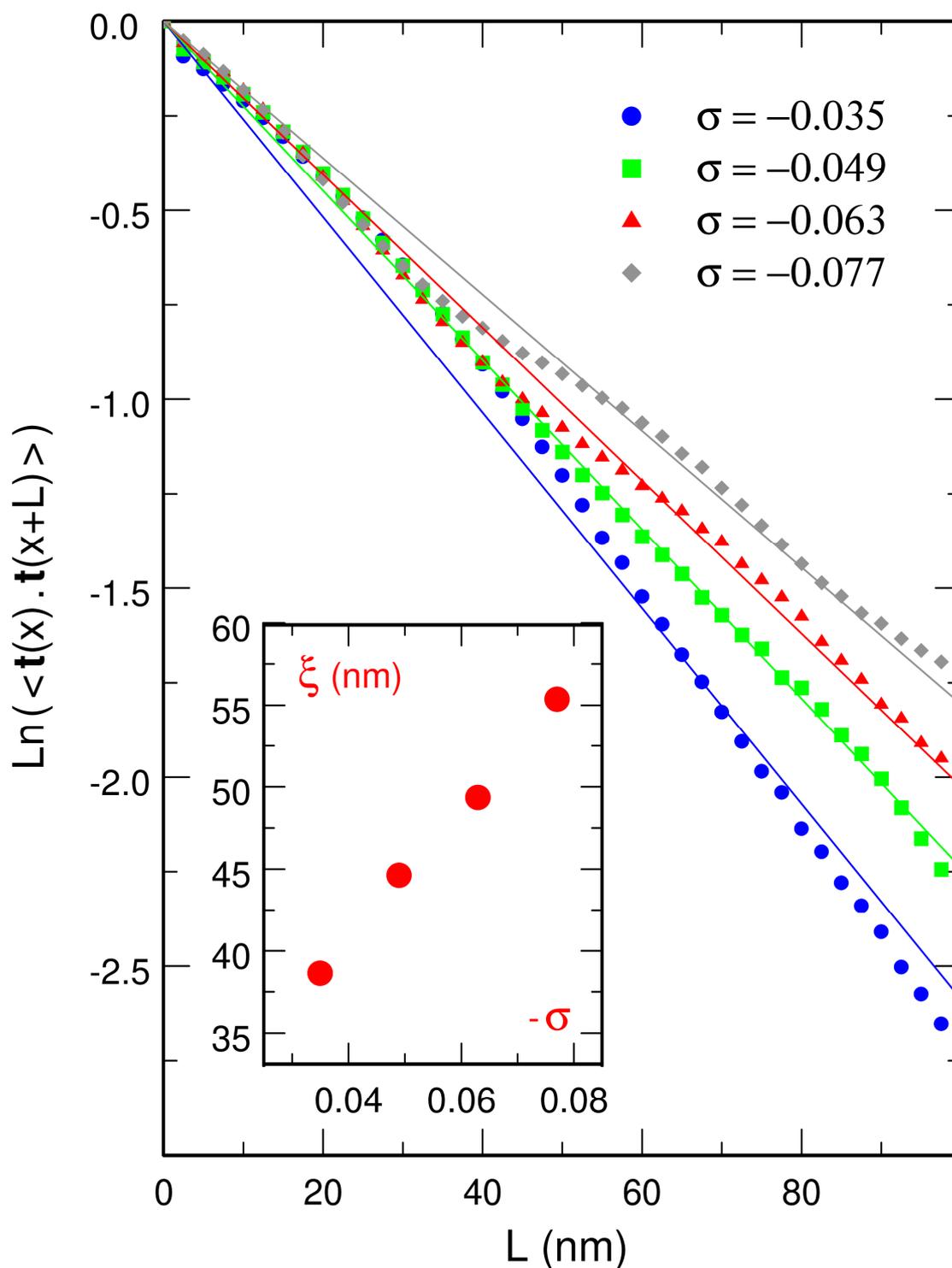


Figure S3 : (Main plot) Plot of the logarithm of the directional correlation function of the segments of the mid curve of plectonemes for different values of the superhelical density σ of a DNA chain with 200 beads (1500 bp). The solid lines show the results of linear fits according to $\text{Ln}(\langle \mathbf{t}(x) \cdot \mathbf{t}(x+L) \rangle) = -L/\xi$, where $\mathbf{t}(x)$ is the tangent to the chain at position x , L the distance between the two segments, and ξ the fitted persistence length (Insert) Evolution of ξ as a function of $-\sigma$.